

REVIEW

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# Isolation and structure elucidation of constituents of *Citrus limon*, *Isodon japonicus*, and *Lansium domesticum* as the cancer prevention agents

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## Abstract

In the course of our research to investigate the cancer prevention potency of natural products derived from plant materials, we isolated fifty-five compounds, including twenty-one new compounds from the peels of *Citrus limon*, aerial parts of *Isodon japonicus*, and leaves of *Lansium domesticum*. The chemical structures of the isolated compounds were elucidated by chemical/physicochemical evidence, and nuclear magnetic resonance spectroscopy and mass spectrometry results. Moreover, the absolute stereochemistry of the new compounds were elucidated by various techniques such as chemical synthesis, modified Mosher's method, Cu-K $\alpha$  X-ray crystallographic analysis, and comparison of experimental and predicted electronic circular dichroism data. The antimutagenic effects of the isolated and structure-elucidated compounds against heterocyclic amines, 3-amino-1,4-dimethyl-5H-pyrido [4,3-*b*]indole (Trp-P-1) and 2-amino-1-methyl-6-phenylimidazo[4,5-*b*]pyridine (PhIP), were evaluated by the Ames test and in vivo micronucleus test. In this review, we present the comprehensive results of the antimutagenic effects of the isolated natural products.

**Keywords:** *Citrus limon*, *Isodon japonicus*, *Lansium domesticum*, Antimutagenic effects, Ames test, Micronucleus test

## Background

Cancer is the leading cause of death worldwide and one of the major risk factors is exposure to the agents that damage the genetic material. These agents are known as genotoxins and, according to their mode of action, are classified into mutagens, carcinogens, or teratogens [1, 2]. Trp-P-1 and PhIP are well known mutagenic and carcinogenic heterocyclic amines that are found in cooked meat. Therefore, it is difficult to completely avoid these risk factors in daily life. On the other hand, previous case-control studies have suggested that the consumption of some plant derived foods such as citrus fruits is associated with a reduced all-cancer incidence [3].

Based on these studies, we searched for antimutagenic materials derived from foods. In the course of this study, we found using the Ames test that the methanolic (MeOH) extracts of *C. limon* [4, 5], aerial parts of *I. japonicus* [6], and leaves of *L. domesticum* [7, 8] showed antimutagenic effects against Trp-P-1 and PhIP. Therefore, we directed our efforts toward the isolation of their constituents and evaluation of the antimutagenic effects of the isolated constituents using the Ames test and in vivo micronucleus test.

## Review

### Compounds obtained from the peels of *C. limon*

The fruits of *Citrus limon* (L.) Burm.f. contain important natural chemical components, such as flavonoids, furanocoumarins, and limonoids [9]. Previous studies have

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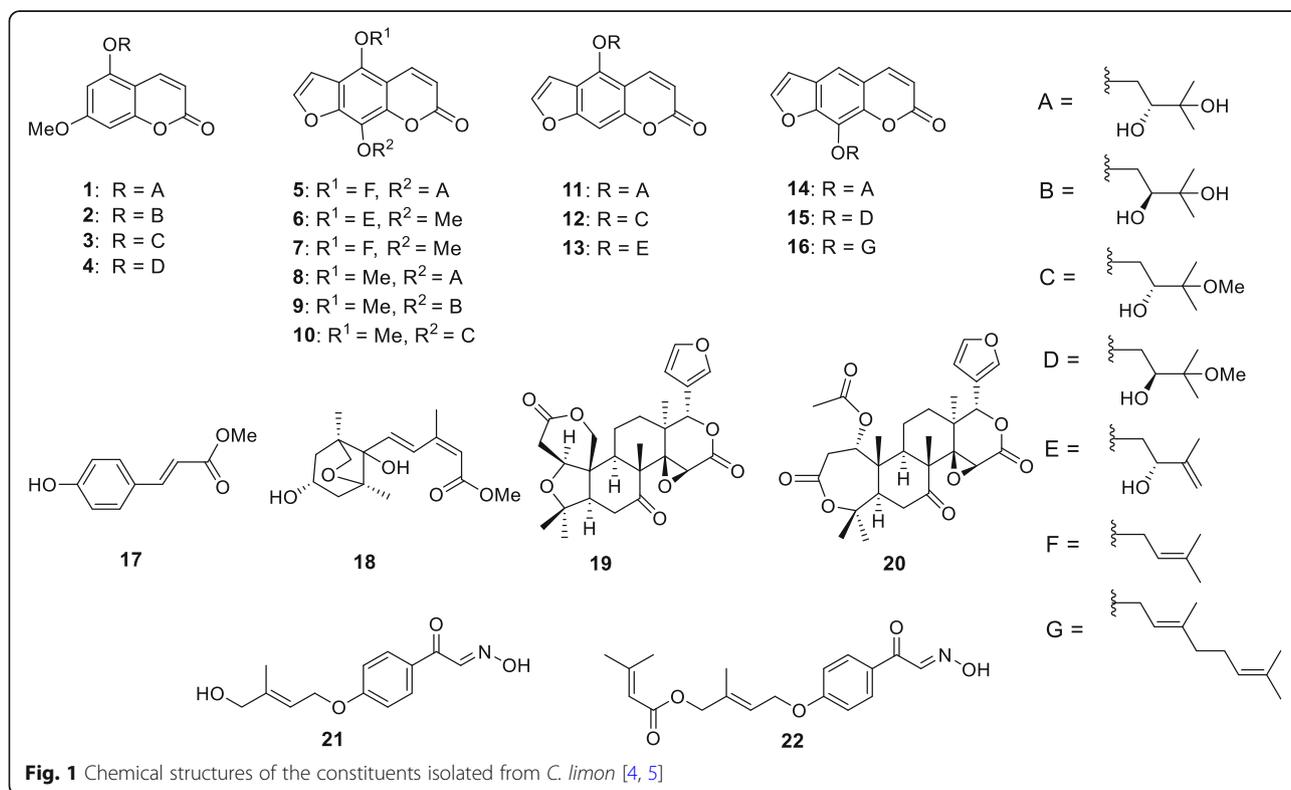
described the biological activities of these compounds, such as the antioxidative, anti-inflammatory, antiallergic, antiviral, antiproliferative, anticarcinogenic, and antimutagenic activities of flavonoids [10], the suppressive effect of limonin on intestinal polyp development in *Apc*-mutant Min mice [11], and the inhibitory effects of furanocoumarins on human CYP 3A4 [12]. In addition, the essential oil of *C. limon* leaf has been shown to act as a central nervous system depressant and anticonvulsant in animal models [11].

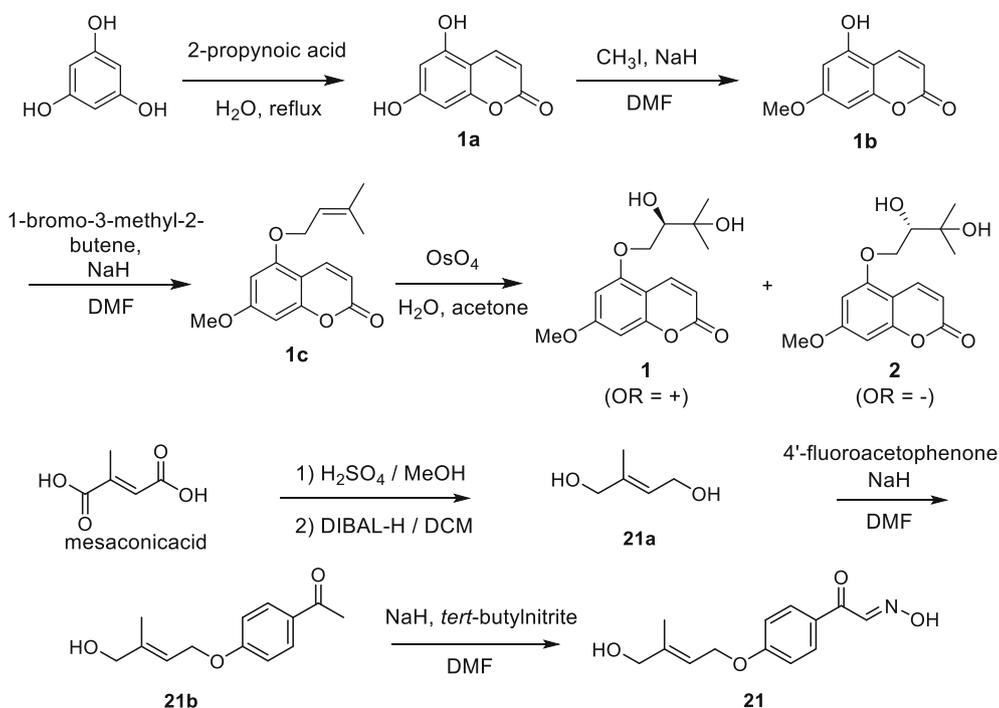
To investigate the chemical structures of the compounds obtained from the peels of *C. limon* and their antimutagenic effects, we started the isolation using the MeOH extract obtained by refluxing the fresh peels. The MeOH extract was partitioned into EtOAc (ethyl acetate)- and H<sub>2</sub>O- soluble fractions and their antimutagenic effects were investigated using the Ames test. The EtOAc- fraction showed the potent antimutagenic effects against Trp-P-1 and PhIP. On the other hand, the H<sub>2</sub>O-soluble fraction showed no detectable effect. Therefore, the EtOAc-soluble fraction was subjected to normal- and reversed-phase column chromatographies, and finally HPLC to isolate four new coumarins, namely, wakayamalimonols A (1), B (2), C (3), and D (4), a new furanocoumarin wakayamalimonol E (5), two new oxime derivatives limonoximes I (21) and II (22), and 11 known furanocoumarins, (+)-apaensin

(6) [13], cnidilin (7) [14], (+)-byakangelicin (8) [15], (-)-byakangelicin (9) [15], (+)-*tert*-O-methylbyakangelicin (10) [16], (+)-oxypeucedanin hydrate (11) [17], (+)-*tert*-O-methyloxypeucedanin hydrate (12) [18], (+)-pangelin (13) [17], (+)-2*a*,3*a*-dihydroxyimperatorin (14) [19], (+)-*O*-methylheraclenol (15) [18], and 8-geranyloxypsoralen (16) [20], a known phenylpropanoid, *p*-coumaric acid methyl ester (17), a known sesquiterpene, 4'-dihydrophaseric acid (18) [21], and two limonoids, limonin (19) [22], and nomilin (20) [23] (Fig. 1). The chemical structures of the isolated compounds were elucidated by chemical and physicochemical evidence, including NMR and MS spectra. The absolute stereochemistries of wakayamalimonol A (1) and B (2) were determined by the modified Mosher's method. In addition, for the identification of the chemical structure and bioassay, wakayamalimonols A (1), B (2) and limonoxime I (21) were synthesized (Scheme 1) [4, 5].

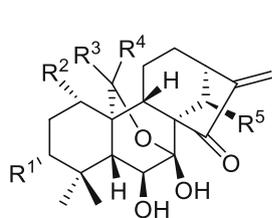
#### Compounds obtained from the aerial parts of *I. japonicus*

*Isodon japonicus* (Burm. f.) *H. Hara* (Lamiaceae) is a perennial plant with a widely distributed in China and Japan [24]. The aerial parts of *I. japonicus* have been used as a traditional herbal medicine for the treatment of gastrointestinal disorders, tumors, and inflammatory diseases [25]. Previous reports have described the structures of diterpenoids [26], flavonoids

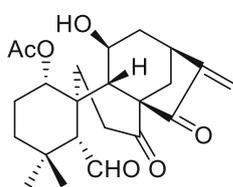




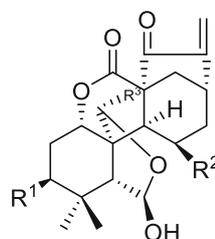
**Scheme 1** Synthesis of wakayamalimonols A (1), B (2) and limonoxime I (21) [4, 5].



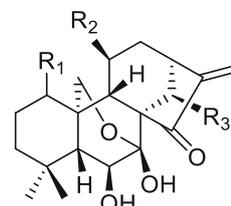
- 23:** R<sup>1</sup> = OH, R<sup>2</sup> = OH, R<sup>3</sup> = H, R<sup>4</sup> = H, R<sup>5</sup> = H  
**25:** R<sup>1</sup> = H, R<sup>2</sup> = OH, R<sup>3</sup> = H, R<sup>4</sup> = H, R<sup>5</sup> = OH  
**26:** R<sup>1</sup> = H, R<sup>2</sup> = OH, R<sup>3</sup> = OH, R<sup>4</sup> = H, R<sup>5</sup> = OH  
**27:** R<sup>1</sup> = H, R<sup>2</sup> = OH, R<sup>3</sup> = H, R<sup>4</sup> = OMe, R<sup>5</sup> = OH  
**28:** R<sup>1</sup> = H, R<sup>2</sup> = OH, R<sup>3</sup> = OMe, R<sup>4</sup> = H, R<sup>5</sup> = OH



**33**

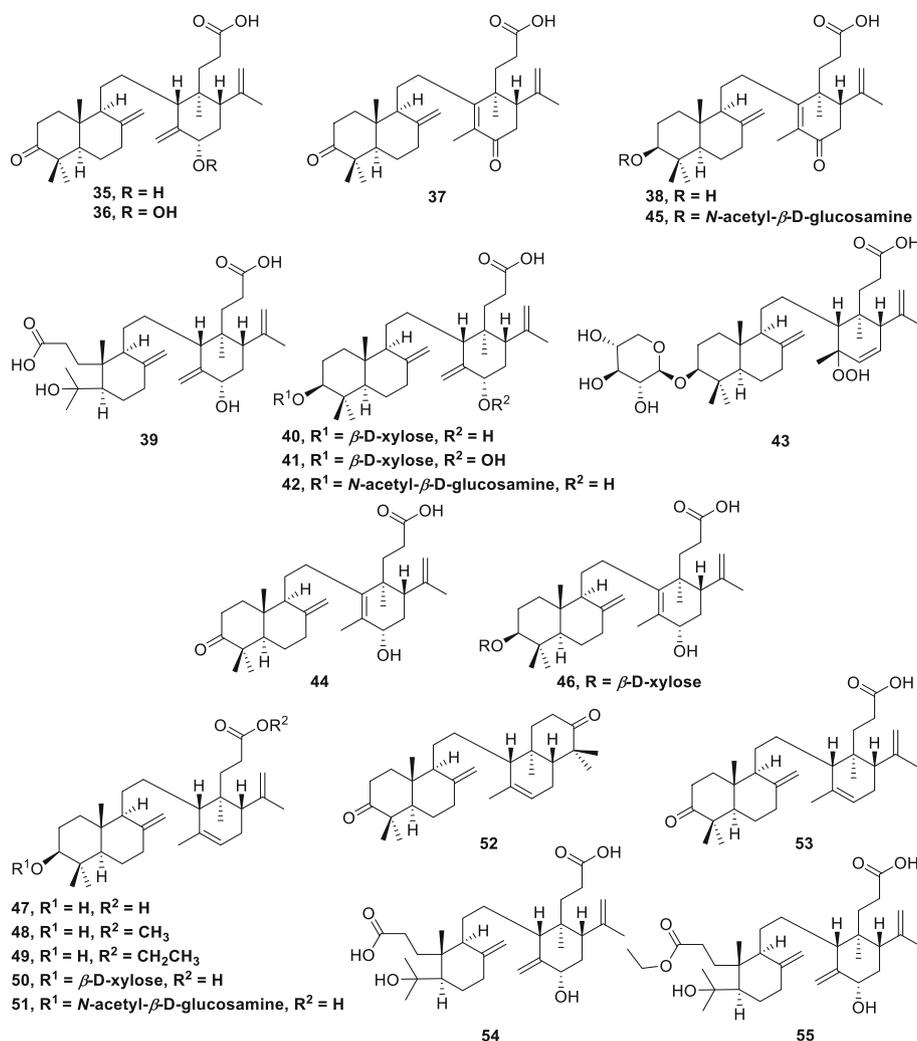


- 29:** R<sup>1</sup> = OH, R<sup>2</sup> = H, R<sup>3</sup> = H  
**30:** R<sup>1</sup> = H, R<sup>2</sup> = OH, R<sup>3</sup> = H  
**31:** R<sup>1</sup> = H, R<sup>2</sup> = H, R<sup>3</sup> = H  
**32:** R<sup>1</sup> = H, R<sup>2</sup> = H, R<sup>3</sup> = OMe



- 24:** R<sup>1</sup> = β-OGlc, R<sup>2</sup> = OH, R<sup>3</sup> = H  
**34:** R<sup>1</sup> = α-OGlc, R<sup>2</sup> = H, R<sup>3</sup> = OH

**Fig. 2** Chemical structures of the constituents isolated from *I. japonicus* [6]



**Fig. 3** Chemical structures of the constituents isolated from *L. domesticum* [7, 8]

[27], and lignans [28] as constituents of *I. japonicus*. Among them, *ent*-kaurane diterpenoids were the most characteristic constituents of *I. japonicus*. Previously, more than forty *ent*-kaurane diterpenoids were identified in phytochemical studies, and some of these compounds have cytotoxic and antibacterial activities [29]. To evaluate the antimutagenic effects of *ent*-kaurane diterpenoids, we first isolated these compounds and elucidated their structures.

From the MeOH extract of the dried aerial parts of *I. japonicus* (cultivated in Tokushima, Japan), two new *ent*-kaurane diterpenoids, named isodoterpenes I (23) and II (24), together with ten known *ent*-kaurane diterpenoids, oridonin (25) [30], hebeirubescensin H (26) [31], rabdternin F (27) [32], rabdternin E (28) [32], enmein (29) [33], nodosin (30) [34], isodocarpin (31) [35], serrin C (32) [36], isodonal (33) [26], and *ent*-7 $\beta$ ,20-epoxy-kaur-16-ene-1 $\beta$ ,6 $\alpha$ ,7 $\alpha$ ,14 $\alpha$ ,15 $\alpha$ -pentanol-1-*O*- $\beta$ -D-glucopyranoside (34) [30], were isolated (Fig. 2). The absolute stereochemistry

of isodoterpene I (23) was elucidated by Cu-K $\alpha$  X-ray crystallographic analysis [6].

#### Compounds obtained from the leaves of *L. domesticum*

*Lansium domesticum* Corr. (Meliaceae), a fruit-bearing tree, grows widely in southeastern Asia [37]. The fruits of this species are edible and are very popular in desserts. Several onoceranoide-type triterpenoids have been isolated from *L. domesticum* peels [38, 39]. Previous reports have described the bioactivities of these onoceranoide-type triterpenoids, such as toxicity against brine shrimp [37], inhibition of leukotriene D4-induced contraction of the guinea pig ileum [40], cytotoxic activity [41], and antibacterial activity against Gram-positive bacteria [42]. However, the bioactivities of these onoceranoide-type triterpenoids have not been investigated thoroughly. Therefore, we attempted to isolate this class of triterpenoids for investigating their biological effects.

**Table 1** Antimutagenic activity of coumarins (**1** and **2**), limonoids (**19** and **20**), oxime and synthetic intermediates (**21**, **21a**, and **21b**), *ent*-kaurane diterpenoids (**25**, **26**, **29**, **30**, and **33**), and onoceranoid-type triterpenoids (**44**, **46–49**, **53**, and **54**) against Trp-P-1 (0.04 µg/plate) using the Ames test [4–8]<sup>a</sup>

	Inhibition % (Based on number of revertant colonies)				
	50 nmol/plate	100 nmol/plate	200 nmol/plate	400 nmol/plate	800 nmol/plate
Coumarins					
<b>1</b> <sup>b</sup>	45.8%	58.7%	64.6%	75.6%	82.3%
<b>2</b> <sup>b</sup>	57.8%	59.8%	67.1%	77.2%	81.4%
Limonoids					
<b>19</b> <sup>b</sup>	10.7%	8.0%	31.0%	46.0%	45.8%
<b>20</b> <sup>b</sup>	38.7%	54.9%	69.8%	83.1%	
Oxime					
<b>21</b> <sup>b</sup>	27.3%	26.2%	40.5%	53.7%	49.0%
Synthetic intermediates of <b>21</b>					
<b>21a</b> <sup>b</sup>	22.2%	20.6%	21.6%	0.2%	–
<b>21b</b> <sup>b</sup>	31.7%	39.2%	51.7%	54.7%	61.4%
<i>ent</i> -Kaurane diterpenoids					
<b>25</b> <sup>b</sup>	34.6%	57.3%	53.7%	65.2%	
<b>26</b> <sup>b</sup>	32.9%	54.4%	60.9%	67.8%	
<b>29</b> <sup>b</sup>	21.0%	47.5%	52.0%	59.9%	
<b>30</b> <sup>b</sup>	46.0%	56.8%	72.8%	73.3%	
<b>33</b> <sup>b</sup>	41.0%	54.9%	68.3%		
Onoceranoid-type triterpenoids					
<b>44</b> <sup>c</sup>	11%	25%	50%	88%	
<b>46</b> <sup>b</sup>	14%	23%	41%	65%	
<b>47</b> <sup>b</sup>	–	73.8%	90.5%	94.2%	
<b>48</b> <sup>b</sup>	0.4%	29.5%	64.8%	77.5%	
<b>49</b> <sup>b</sup>	3.7%	7.9%	8.7%	42.6%	
<b>53</b> <sup>b</sup>	5.1%	79.9%	91.5%	92.4%	
<b>54</b> <sup>c</sup>	–	25.0%	61.0%	93.0%	

<sup>a</sup> Nobiletin was used as a reference compound. It showed 56% inhibition at 20 nmol/plate<sup>b</sup> Compounds showed no antibacterial activity [< 5.0% inhibition at highest concentration] against the *S. typhimurium* TA98 strain<sup>c</sup> Compounds showed weak antibacterial activity [**44**: 23.2% inhibition at 400 nmol/plate, **54**: 23.0% inhibition at 400 nmol/plate] against the *S. typhimurium* TA98 strain

From the MeOH extract of *Lansium domesticum* dried leaves, twelve new compounds, namely, lansium acids I (**35**), II (**36**), III (**37**), IV (**38**), V (**39**), VI (**40**), VII (**41**), VIII (**42**), IX (**43**), X (**44**), XI (**45**), and XII (**46**) together with nine known compounds, lansiolic acid (**47**) [40], methyl lansiolate

(**48**) [40], ethyl lansiolate (**49**) [42], lansioside C (**50**) [40], lansioside B (**51**) [40], 8,14-*seco*-gammacera-7,14-diene-3,21-dione (**52**) [43], lansionic acid (**53**) [37], lansic acid (**54**) [37], and lamesticum A (**55**) [42] were isolated (Fig. 3). The absolute stereo structures of the new compounds were

**Table 2** Antimutagenic activity of furanocoumarins (**11**, **12**, **14**, and **15**) against Trp-P-1 (0.04 µg/plate) using the Ames test [4]<sup>a</sup>

	Inhibition % (Based on number of revertant colonies)				
	3.1 nmol/plate	6.3 nmol/plate	12.5 nmol/plate	25 nmol/plate	50 nmol/plate
Furanocoumarins					
<b>11</b>	22%	13.9%	25.0%	31.5%	40.9%
<b>12</b>	27.0%	35.9%	51.6%	59.5%	74.7%
<b>14</b>	25.0%	40.5%	49.7%	62.8%	73.7%
<b>15</b>	5.6%	5.6%	18.1%	36.6%	51.2%

<sup>a</sup> No compound showed antibacterial activity [< 5.0% inhibition at highest concentration] against the *S. typhimurium* TA98 strain at the tested concentrations

**Table 3** Antimutagenic activity of coumarins (**1** and **2**), limonoids (**19** and **20**), oxime and synthetic intermediates (**21**, **21a**, and **21b**), *ent*-kaurane diterpenoids (**25**, **26**, **29**, **30**, and **33**), and onoceranoid-type triterpenoids (**47–49**, **53**, and **54**) against PhIP (1.0 µg/plate) using the Ames test [4–8]<sup>a,b</sup>

	Inhibition % (Based on number of revertant colonies)				
	50 nmol/plate	100 nmol/plate	200 nmol/plate	400 nmol/plate	800 nmol/plate
Coumarins					
<b>1</b> <sup>b</sup>	17.5%	29.1%	48.4%	48.6%	81.4%
<b>2</b> <sup>b</sup>	18.2%	35.4%	62.2%	77.6%	86.3%
Limonoids					
<b>19</b> <sup>b</sup>	27.0%	39.0%	25.5%	50.3%	46.7%
<b>20</b> <sup>b</sup>	23.6%	44.2%	74.9%	79.8%	
Oxime					
<b>21</b> <sup>b</sup>	12.2%	11.1%	29.7%	47.1%	63.7%
Synthetic intermediates of <b>21</b>					
<b>21a</b> <sup>b</sup>	8.6%	7.4%	0.7%	–	–
<b>21b</b> <sup>b</sup>	10.7%	18.0%	39.0%	53.3%	68.1%
<i>ent</i> -Kaurane diterpenoids					
<b>25</b> <sup>b</sup>	55.8%	62.8%	73.7%	80.7%	
<b>26</b> <sup>b</sup>	54.3%	72.0%	79.5%	85.5%	
<b>29</b> <sup>b</sup>	24.0%	43.2%	62.3%	74.2%	
<b>30</b> <sup>b</sup>	51.0%	64.8%	81.4%	85.5%	
<b>33</b> <sup>b</sup>	71.0%	79.5%	80.4%		
Onoceranoid-type triterpenoids					
<b>47</b> <sup>b</sup>	30.8%	72.7%	93.7%	94.9%	
<b>48</b> <sup>b</sup>	14.3%	33.7%	54.3%	74.4%	
<b>49</b> <sup>b</sup>	4.1%	17.2%	22.5%	41.0%	
<b>53</b> <sup>b</sup>	36.8%	65.2%	94.1%	93.9%	
<b>54</b> <sup>c</sup>	18.6%	32.4%	63.2%	95.4%	

<sup>a</sup> Nobiletin was used as a reference compound. It showed 56% inhibition at 20 nmol/plate

<sup>b</sup> Compounds showed no antibacterial activity [< 5.0% inhibition at highest concentration] against the *S. typhimurium* TA98 strain

<sup>c</sup> Compound showed weak antibacterial activity [**54**: 23.0% inhibition at 400 nmol/plate] against the *S. typhimurium* TA98 strain

established by comparison of the experimental and predicted electronic circular dichroism data [7, 8].

#### Evaluation of the antimutagenic effects of isolated compounds using the Ames test

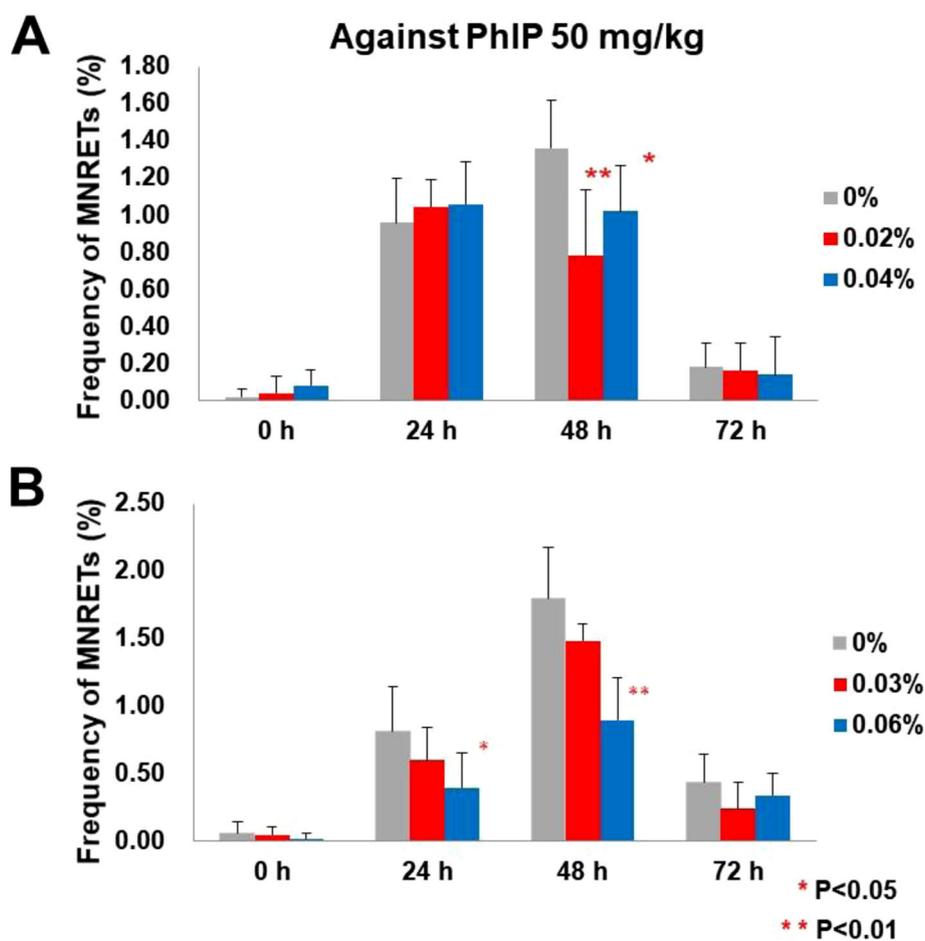
The antimutagenic effects of the isolated compounds were evaluated against Trp-P-1 and PhIP by the Ames test using the *S. typhimurium* TA98

strain (Tables 1, 2, 3 and 4). Trp-P-1 and PhIP are well known mutagenic and carcinogenic heterocyclic amines found in cooked meat. We used nobiletin as the positive control that have been reported to have antimutagenic effects using the Ames test [44]. As shown in Tables 2 and 4, among the compounds isolated from the peels of *C. limon*, furanocoumarins, (+)-*tert*-*O*-methoxyxypeucedanin hydrate

**Table 4** Antimutagenic activity of furanocoumarins (**11**, **12**, **14**, and **15**) against PhIP (1.0 µg/plate) using the Ames test [4]<sup>a</sup>

	Inhibition % (Based on number of revertant colonies)				
	3.1 nmol/plate	6.3 nmol/plate	12.5 nmol/plate	25 nmol/plate	50 nmol/plate
Furanocoumarins					
<b>11</b>	22.0%	28.0%	29.7%	46.3%	64.0%
<b>12</b>	47.0%	57.7%	68.5%	73.4%	83.4%
<b>14</b>	34.0%	41.7%	51.7%	67.6%	70.1%
<b>15</b>	47.1%	62.1%	65.3%	69.6%	78.7%

<sup>a</sup> No compound showed antibacterial activity [< 5.0% inhibition at highest concentration] against the *S. typhimurium* TA98 strain at the tested concentrations



**Fig. 4** Frequency of MNRETs from peripheral blood of mice treated with mutagen {PhIP (50 mg/kg bw)}. Significant difference: \* $0.01 < P < 0.05$ ; \*\* $P < 0.01$  (Student's *t* test). Each point represents the mean and standard deviation of five mice. **a** Normal or sample feeds that included limonin (19) at low or high dose (0.02% or 0.04%, w/w) were given ad libitum. **b** Normal or sample feeds that included lansionic acid (53) at low or high dose (0.03% or 0.06%, w/w) were given ad libitum [4, 8]

(12) [inhibition against Trp-P-1: 74.7% at 50 nmol/plate] and (+)-2*a*,3*a*-dihydroxyimperatorin (14) [inhibition against Trp-P-1: 73.7% at 50 nmol/plate] showed potent antimutagenic effects. The antimutagenic effects of 12 and 14 were much stronger than that of the reference compound, nobiletin [inhibition against Trp-P-1: 56.0% at 80 nmol/plate]. On the other hand, coumarins (1 and 2), and limonoids (19 and 20) showed weak but dose-dependent antimutagenic effects against Trp-P-1 and PhIP. The oxime (21) and synthetic intermediate (21b) showed antimutagenic effect, but the synthetic intermediate without phenyl group (21a) did not show any effects. These results suggest that a phenyl group is necessary for the antimutagenic effects of this class of compounds [4, 5]. The *ent*-kaurane diterpenoids (26, 27, 30, 31, and 34) from *I. japonicus* also showed antimutagenic effects against Trp-P-1 [inhibition against Trp-P-1: 47.4–

57.3% at 100 nmol/plate]. While their effects were not stronger than that of nobiletin, our study represented the first evaluation of the antimutagenic activities of *ent*-kaurane diterpenes [6]. Among the isolated onoceranooid-type triterpenoids, the antimutagenic effects of lansiolonic acid (47) [inhibition against Trp-P-1: 73.8% at 100 nmol/plate] and lansionic acid (53) [inhibition against Trp-P-1: 79.9% at 100 nmol/plate] against Trp-P-1 were equivalent to that of nobiletin. In addition, interesting structure-activity relationships were suggested. Specifically, the compound with a carboxylic acid moiety (47) showed more potent antimutagenic effects than its analogs with either a methyl ester moiety (48) [inhibition against Trp-P-1: 29.5% at 100 nmol/plate] or ethyl ester moiety (49) [inhibition against Trp-P-1: 7.9% at 100 nmol/plate]. Among the esters, the ethyl ester compound 49 showed weaker effects than the methyl ester 48 [7, 8]. The antibacterial

activity of tested compounds against the *S. typhimurium* TA98 strain were tested using nutrient agar containing NaCl and all compounds showed weak [44: 23.2% inhibition at 400 nmol/plate, 54: 23.0% inhibition at 400 nmol/plate] or no (other compounds: < 5.0% inhibition at highest concentration in Table 1) antibacterial activities [4–8].

### Evaluation of antimutagenic effects of limonin (19) and lansionic acid (53) using in vivo micronucleus test

To examine the antimutagenic effects of limonin (19) and lansionic acid (53) in vivo, we conducted a micronucleus test using the peripheral blood of male ICR mice. These two compounds were isolated enough amount and are the major constituents of *C. limon* and *L. domesticum*. The micronucleus test detects chromosomal damage induced by genotoxic/carcinogenic compounds, and it has been used to evaluate antimutagenic agents in vivo. We gave either normal feed or sample feed that included the limonin (19) or lansionic acid (53) at low or high dose. The mouse tail vein blood (5  $\mu$ L) was taken prior to the administration of PhIP, and at 24, 48, and 72 h after administration. As a result, limonin (19) and lansionic acid (53) significantly decreased the frequency of micronucleated reticulocytes (MNRETs) treated with PhIP at 24 h and 48 h after the administration (Fig. 4) [4, 7].

### Conclusions

In conclusion, twenty-one new compounds and thirty-four known compounds including coumarins, furanocoumarins, oximes, *ent*-kaurane diterpenoids, and onoceranoid-type triterpenoids were isolated from the peels of *C. limon*, aerial parts of *I. japonicus*, and leaves of *L. domesticum*. Among them, furanocoumarins showed the strongest antimutagenic effects in the Ames test. It is important to note that furanocoumarins were well known Cytochrome P450 (CYP) 1A2 inhibitors [45], and the mutagenicity of Trp-P-1 and PhIP depends on their bioactivation by CYP 1A2 [46]. These facts suggest that one of the mechanisms of the antimutagenic effects of furanocoumarins against Trp-P-1 and PhIP may be the inhibition of the CYP 1A2 enzyme. In addition, the oral intake of limonin (19) and lansionic acid (53), which are the major constituents of *C. limon* and *L. domesticum*, respectively, showed significant antimutagenic effects against PhIP in the in vivo micronucleus test. These results suggest that these compounds and plant materials are likely useful agents for cancer prevention.

### Abbreviations

ECD: Electronic circular dichroism; HPLC: high-performance liquid chromatography; ICR: Institute of Cancer Research; MNRET: micronucleated reticulocyte; NMR: nuclear magnetic resonance; Trp-P-1: 3-amino-1,4-

dimethyl-5*H*-pyrido[4,3-*b*]indole; PhIP: 2-amino-1-methyl-6-phenylimidazo[4,5-*b*]pyridine

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### Authors' contributions

The author(s) read and approved the final manuscript.

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### Availability of data and materials

Not applicable.

### Ethics approval and consent to participate

Not applicable.

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All authors have read the manuscript and approved its submission and publication.

### Competing interests

The authors declare that they have no competing interests.

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